

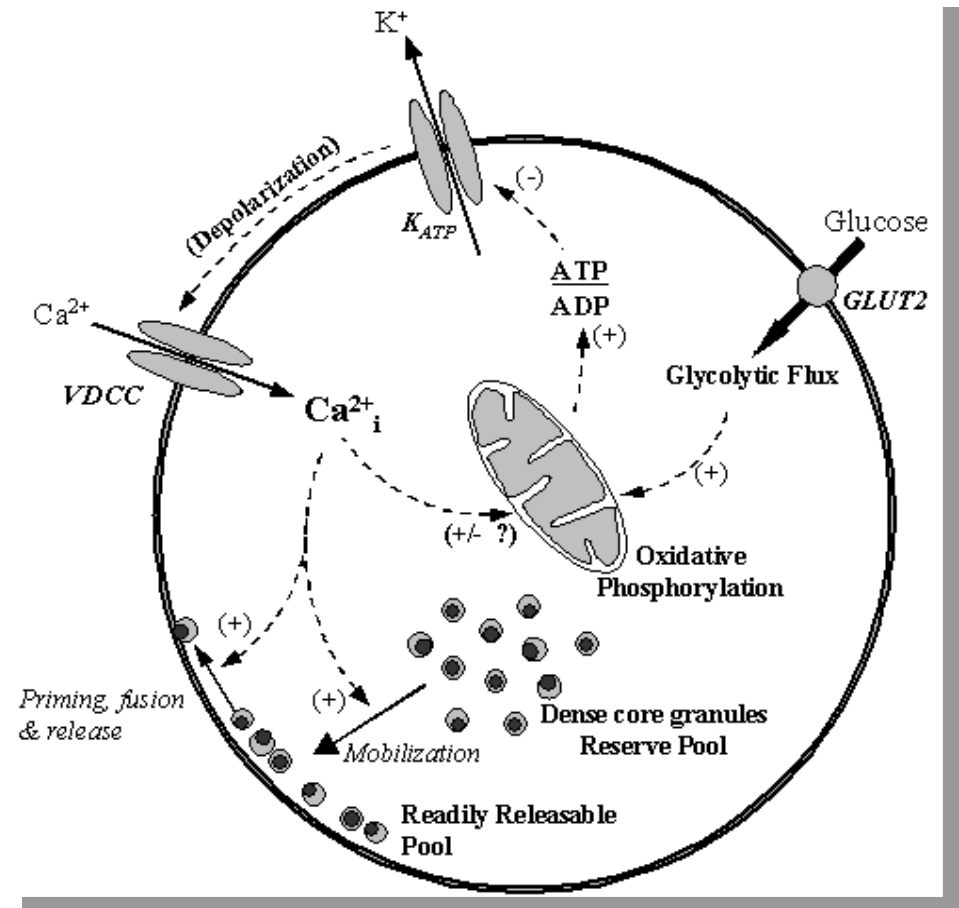
Spanning time scales and levels of organization of insulin secretion with mathematical modeling: From seconds to hours, from molecules to organ

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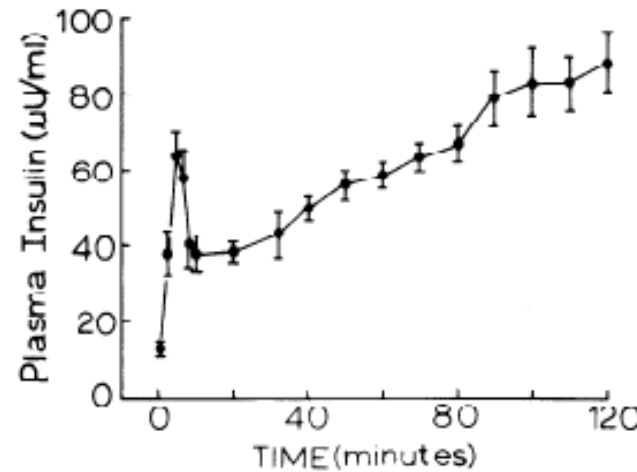
Glucose stimulated insulin secretion

- Insulin is secreted from the pancreatic beta-cells in response to (mainly) glucose
- Insulin is stored in *secretory granules*
- Released by Ca^{2+} *triggered exocytosis*

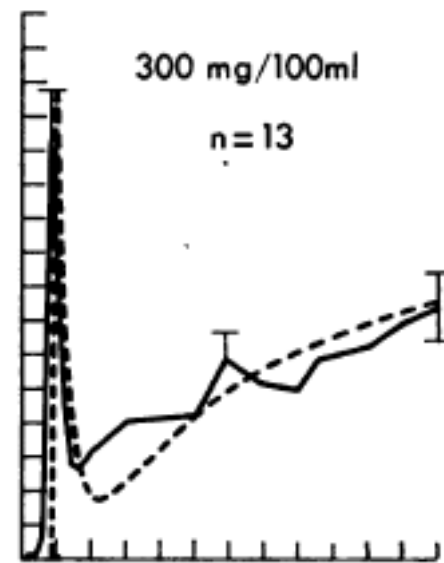


Phasic insulin secretion

- Insulin is secreted in typical biphasic pattern in response to glucose-step
- Seen *in vivo*, from pancreases and from islets
- Pools of granules?



In vivo glucose clamp
(De Fronzo et al., 1979)



Perfused rat pancreas
(Grotsky, 1972)

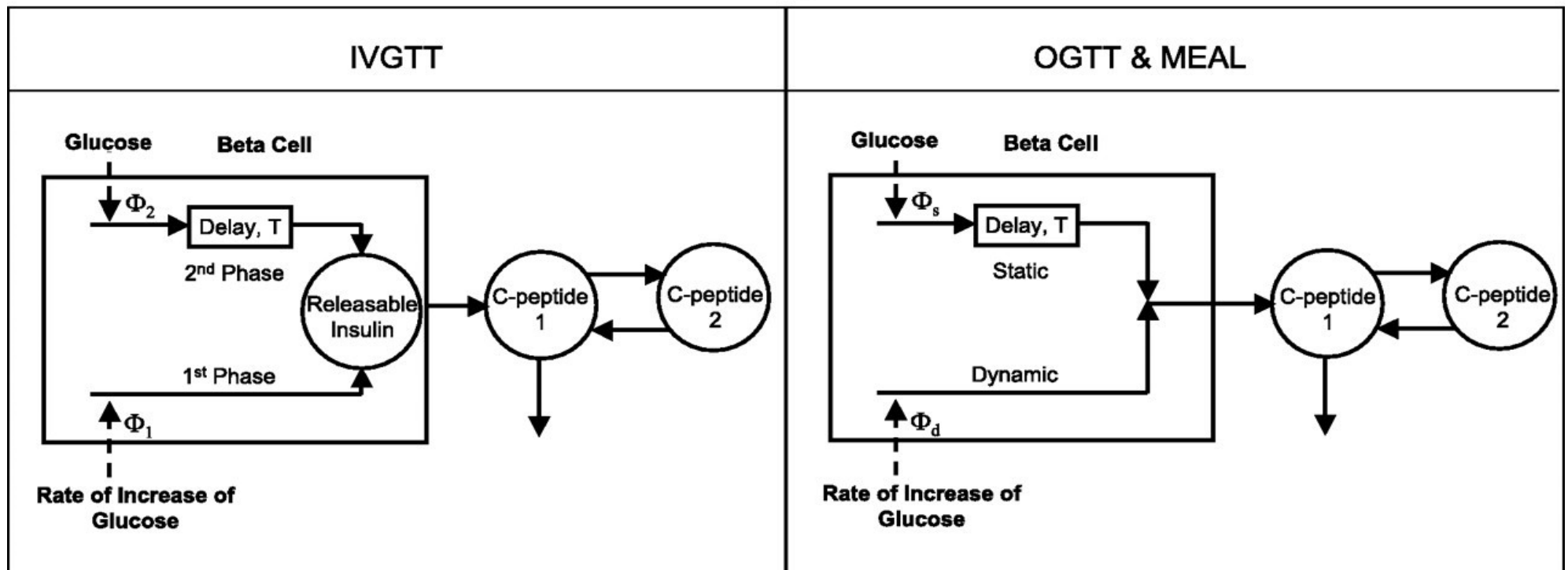
Minimal models of insulin secretion

– estimated from in vivo data

- C-peptide minimal model allows estimation of dynamic and static beta-cell responsiveness

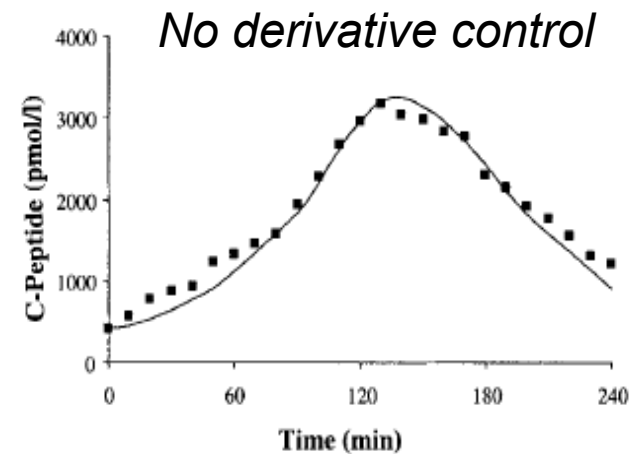
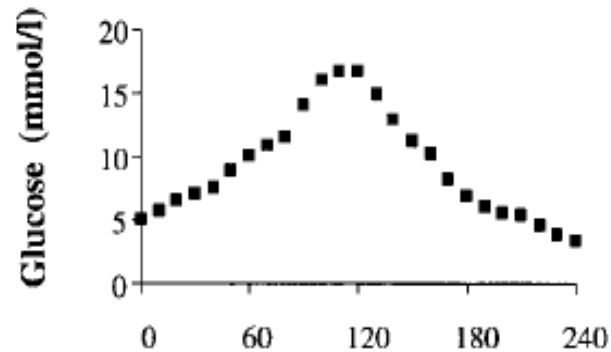
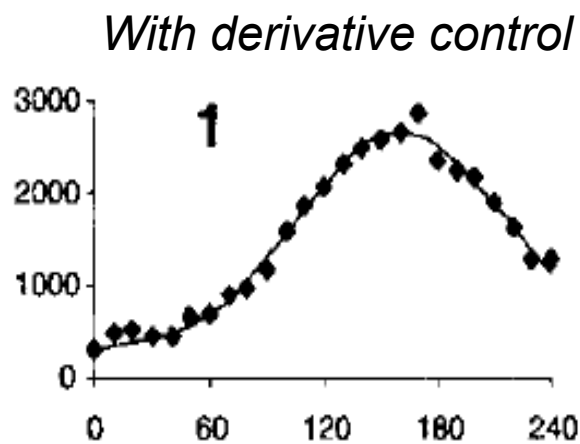
(Toffolo et al., Diabetes 1995, AJP 2001; Cobelli et al., AJP 2007)

Beta Cell Responsivity Minimal Models



Dynamic secretion term

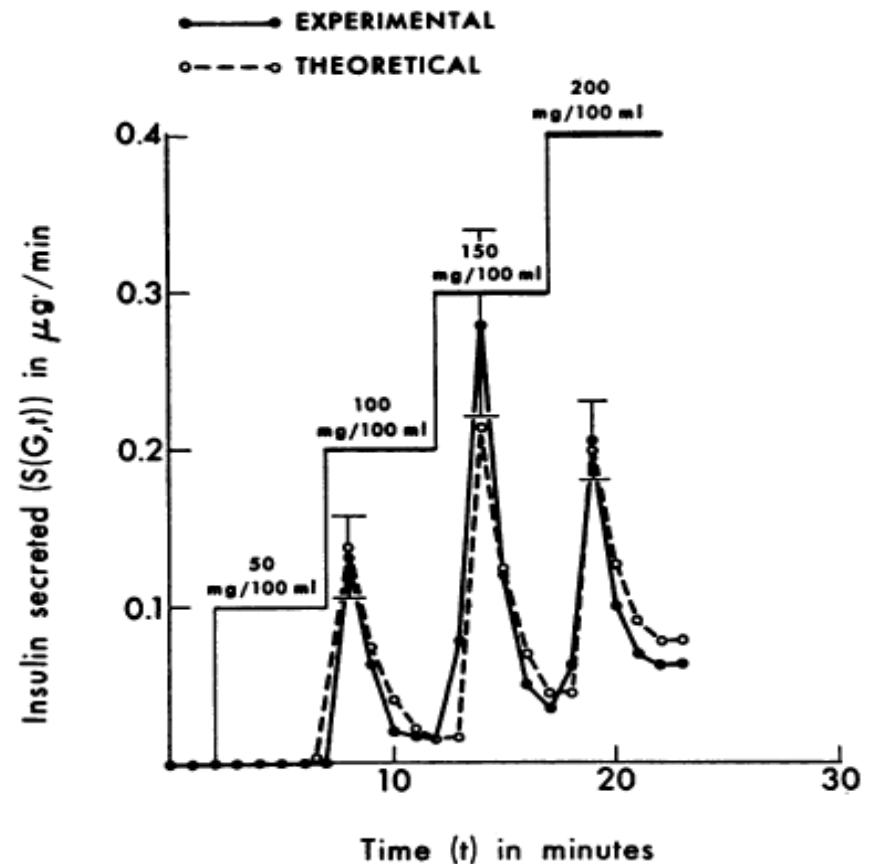
- Derivative control: The pancreas responds not only to glucose concentration G (“static”, with delay), but also to rate-of-change dG/dt
- Necessary to fit in vivo data
(Graded up-down: Toffolo et al., AJP 2001; OGTT: Breda et al., Diabetes 2001)



- Where does it come from?

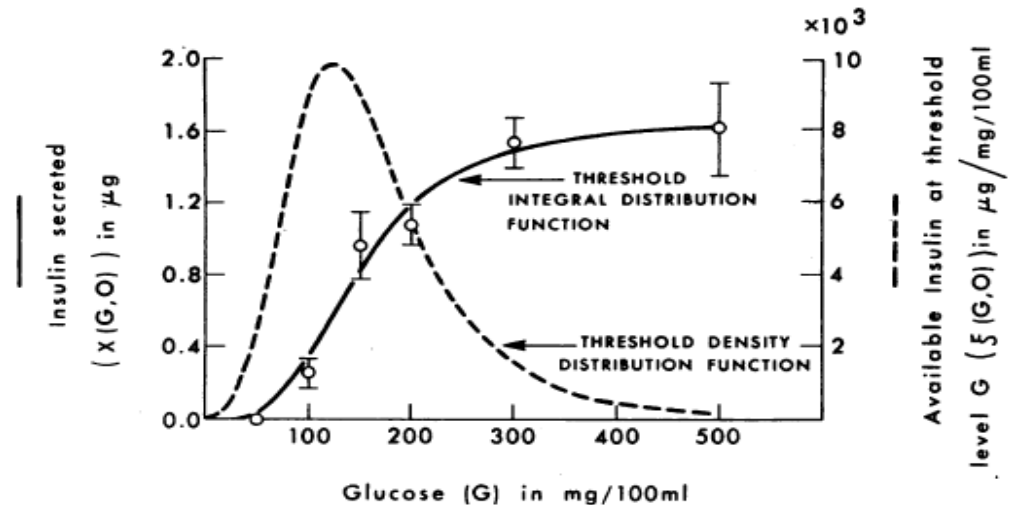
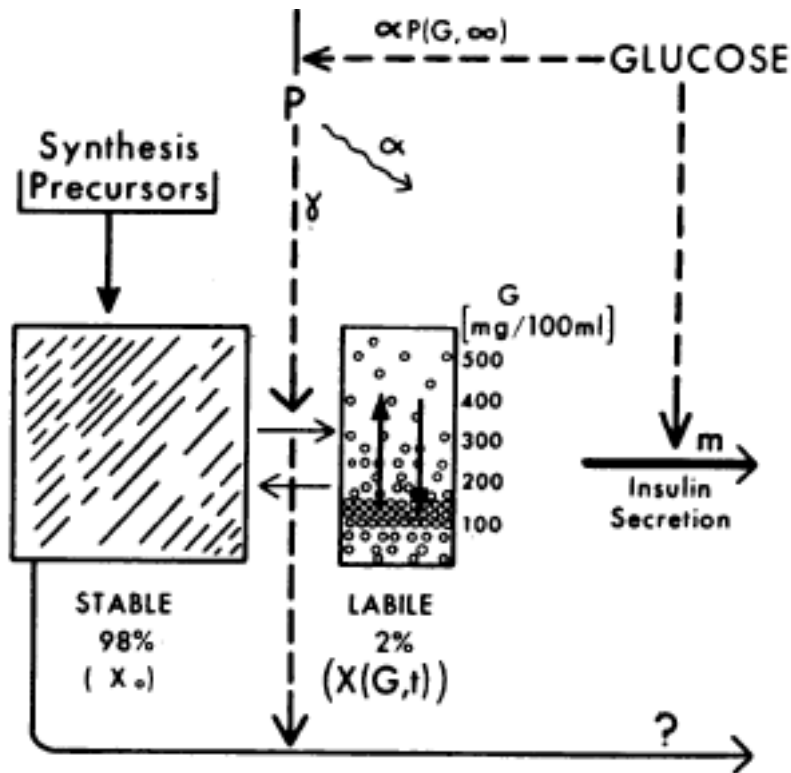
Staircase experiment

- Rat pancreas
(Grotsky, JCI 1972)
- Sum of peaks =
peak at max
concentration →
threshold hypothesis
(Grotsky, JCI 1972)



Grodsky's threshold hypothesis

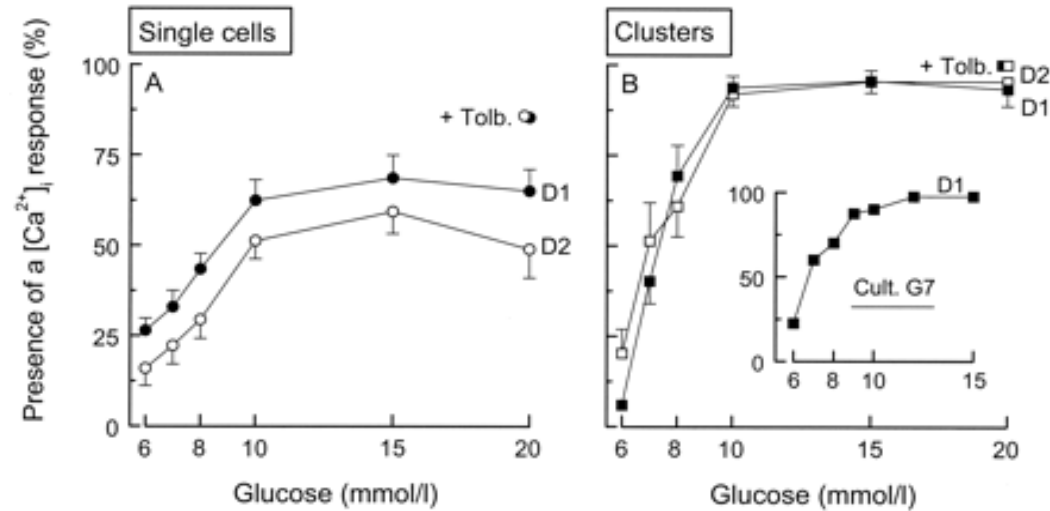
- Two pools of “packages”
- “Labile packages” are heterogeneous, different glucose thresholds for release



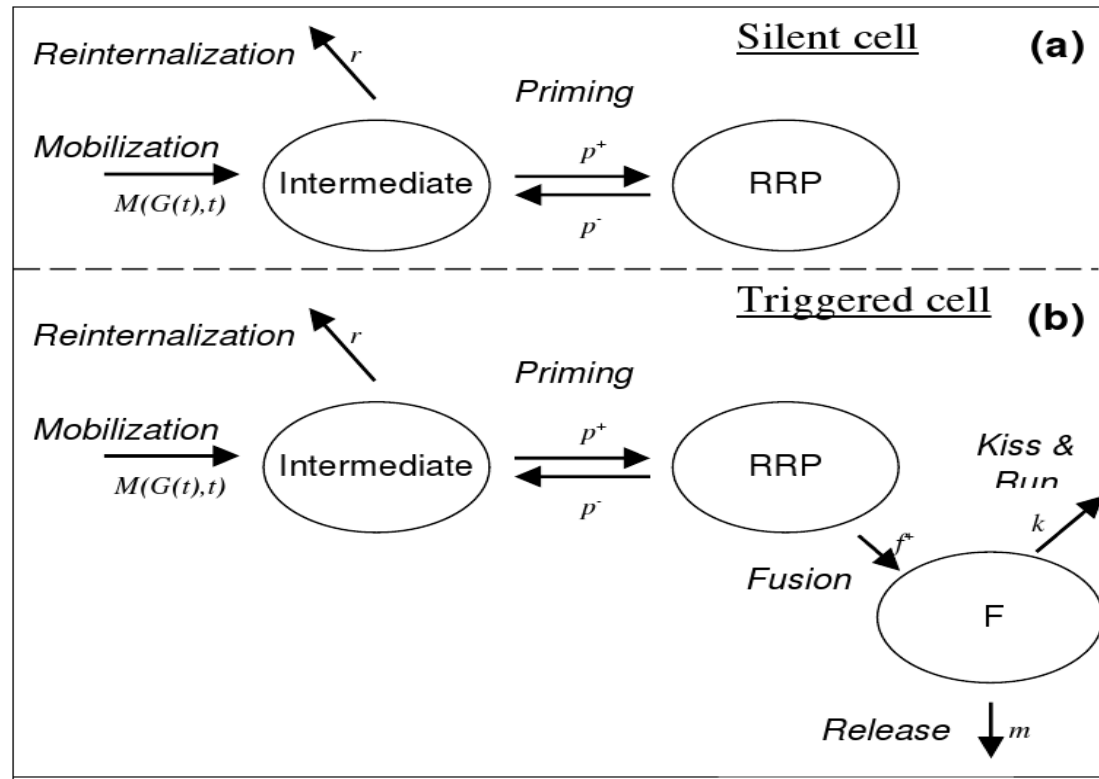
Model development

- Previous attempts:
Grotsky (JCI '72),
Landahl & Grotsky (BMB '82)
- Recent granule models:
Bertuzzi et al. (AJP '07),
Chen et al. (BJ '08),
Pedersen & Sherman
(PNAS '09)
- Cells (not granules!) activate
at different glucose
concentrations →
heterogeneous RRP

(Pedersen et al.,
Phil Trans Roy Soc A 2008)

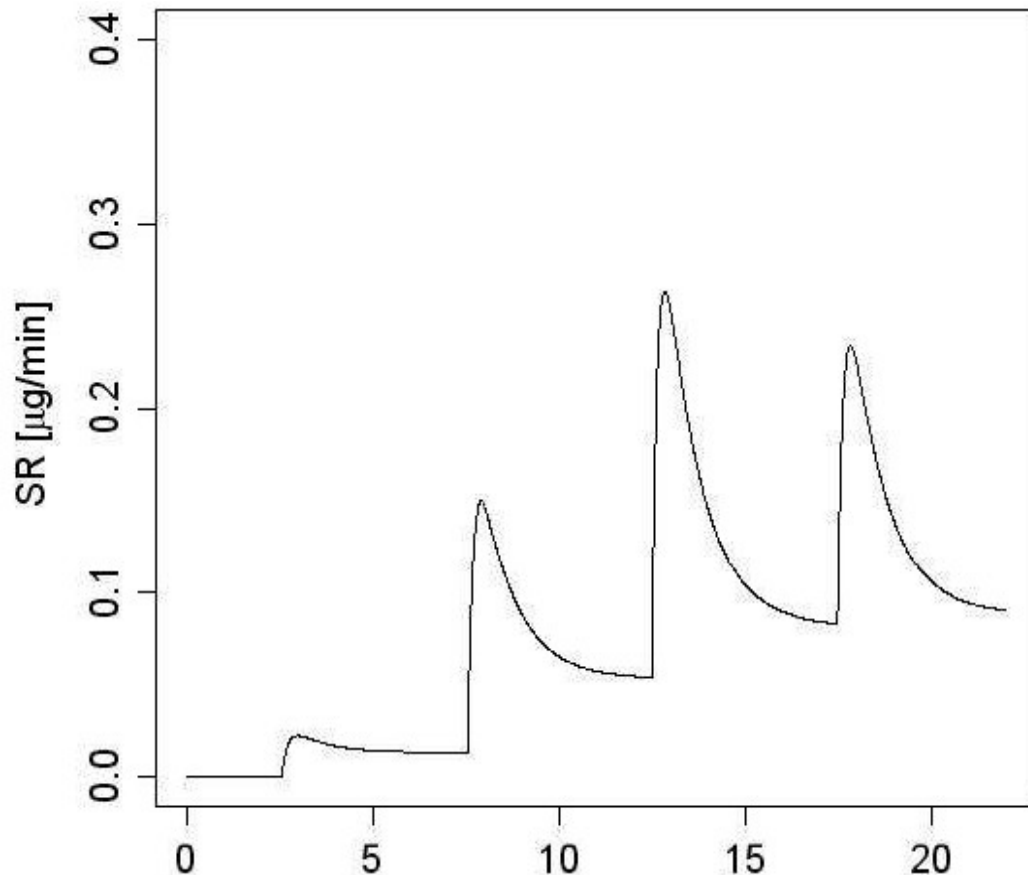


(Jonkers & Henquin, Diabetes 2001)



It does the job...

Staircase



- Other recent models do not reproduce staircase
- Heterogeneous RRP allows reproduction of staircase experiment
- In contrast to Grodsky, due not to threshold on *granules* but on *cells* (or islets?) as seen in experiments

Where does derivative control come from?

(Pedersen et al., AJP 2010)

- Threshold distribution underlies derivative control (Grotsky, JCI 1972; Licko, Bull Math Biol 1973)

- Here:

- $SR = mF$

- $dF/dt = -(m+k)F + fH(G), \quad H(G) = \int_0^G h(g)dg$

- $dH/dt = \int_0^G dh(g)/dt dg + h(G) dG/dt$
 $= - (f+p^-)H(G) + p^+ I \Phi(G) + h(G) dG/dt$

- Assume quasi steady-state

- $SR(t) = \text{const} [p^+ I(t, \tau) \Phi(G(t)) + h(G(t)) dG/dt(t)]$

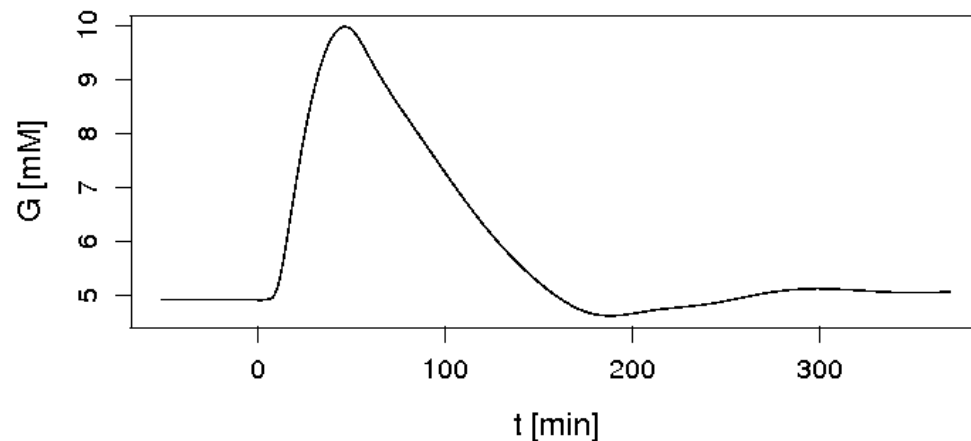
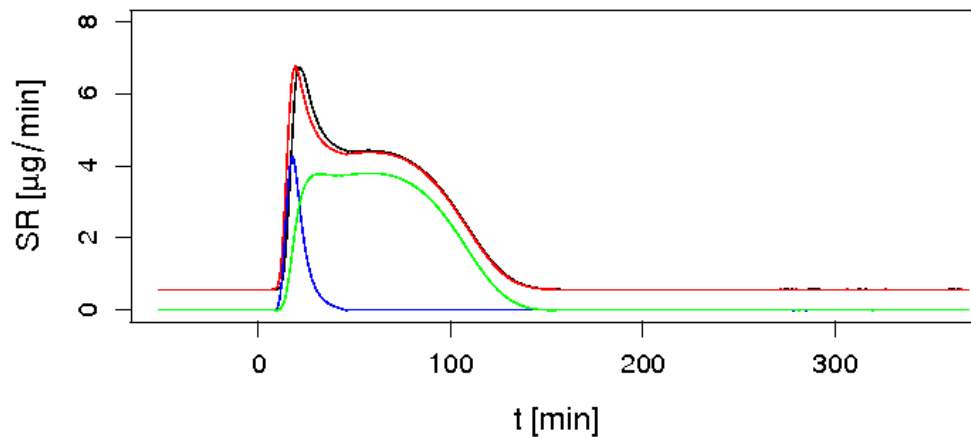


Static responsivity



Dynamic responsivity

Relative contributions of dynamic vs. static secretion



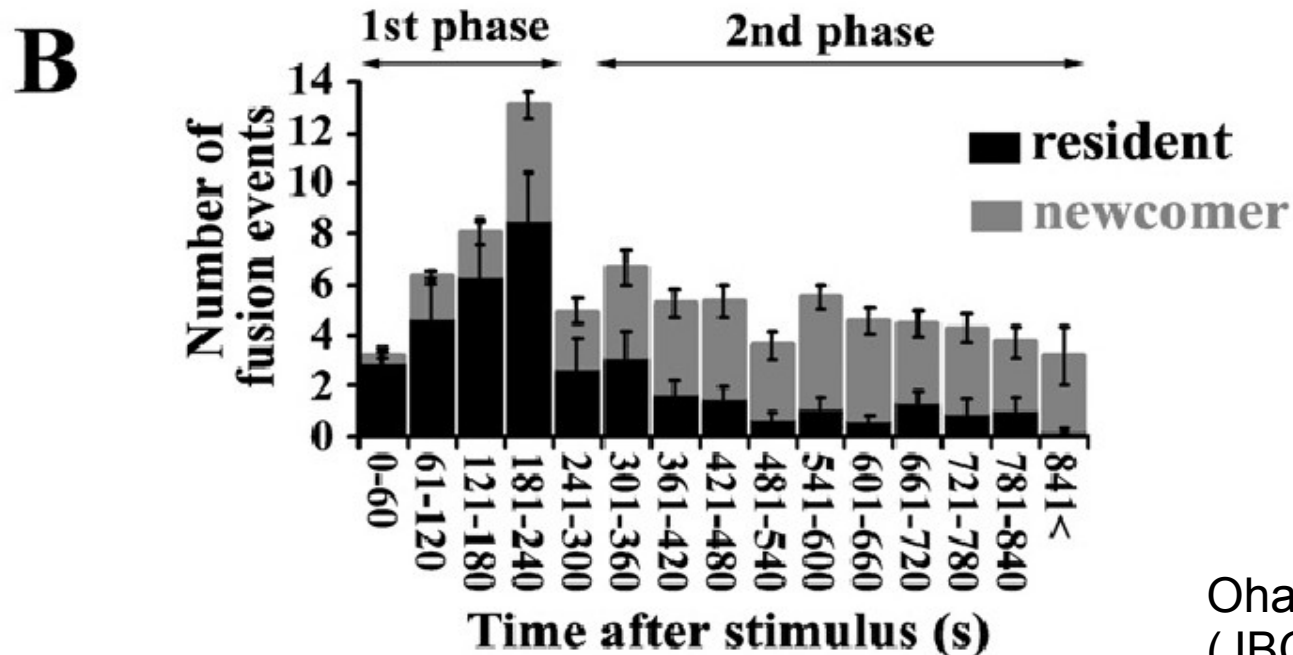
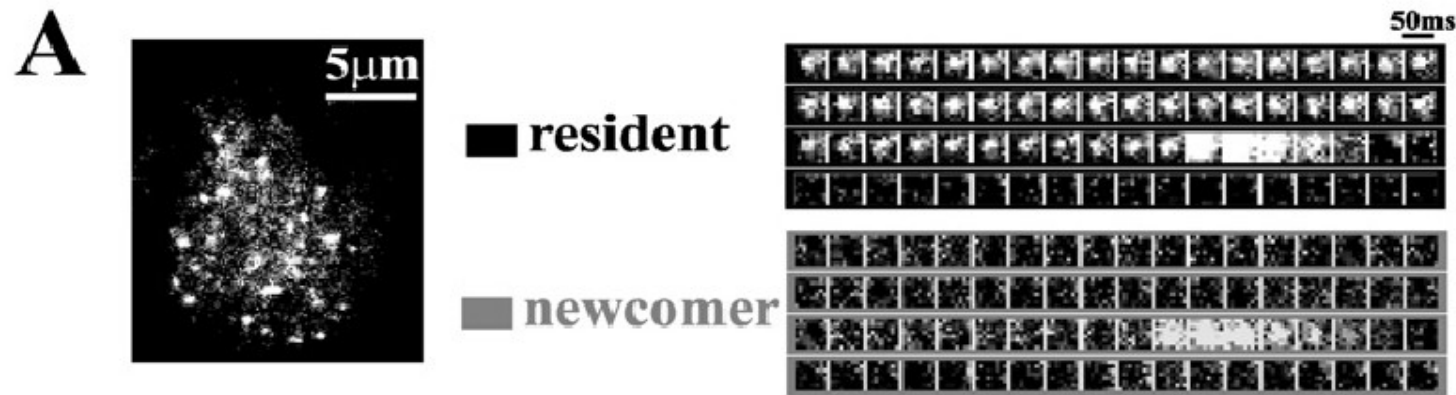
- Glucose profile following a meal
- Model parameters adjusted to give reasonable C-peptide data
- Legend:
 - ♦ Full model
 - ♦ Approximation
 - ♦ Dynamic
 - ♦ Static

Conclusions (part 1)

- Relatively simple model, but founded on biologically established principles (non-phenomenological)
- Can explain static and dynamic secretion terms
 - *Dynamic* due to recruitment of cells (or islets?)
 - *Static* due to refilling of RRP (introduces delay)
- The model could (should!?) be coupled to models of calcium dynamics
- Such models provide mechanistic underpinning of the assumptions of the minimal models

Granules → *cells* → *pancreas*
- ... and could help in interpreting in vivo data (disturbances in diabetics?)

Distinct mechanisms account for 1st and 2nd phase secretion

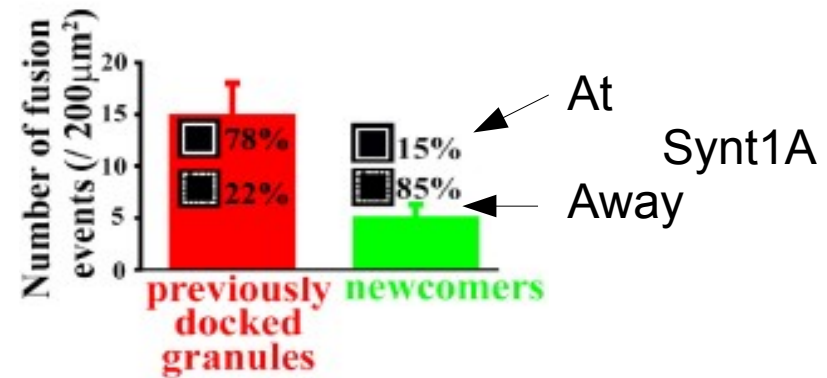


1st (resp. 2nd) phase secretion occurs mainly near (resp. away from) Ca²⁺ channels

Docked granules fuse *at* Synt1A clusters (~80%), newcomers fuse *away from* Synt1A clusters (~85%)

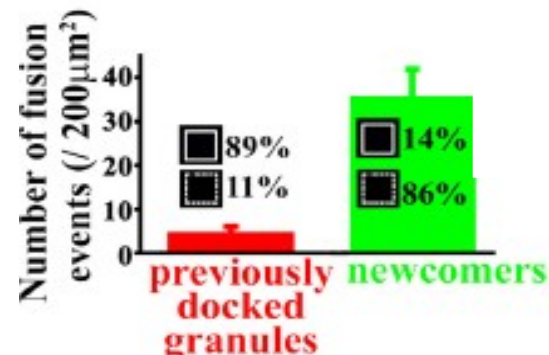
Synt1A clusters are co-located with L-type Ca²⁺ channels (Yang et al., PNAS 1999)

1st



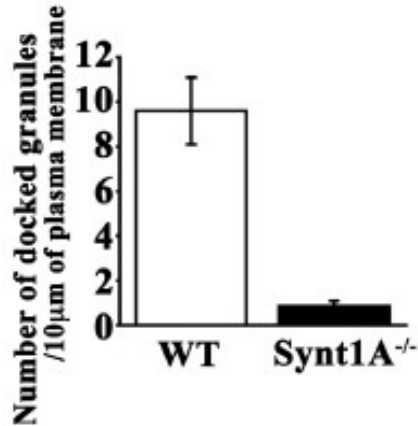
Ohara-Imaizumi et al. (J Cell Biol 2007)

2nd



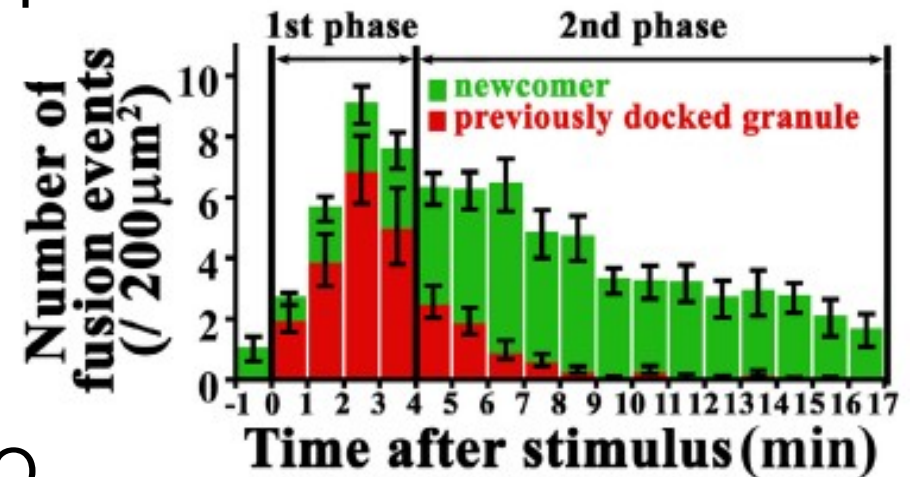
... and docking is not a prerequisite for the 2nd phase

Syntaxin (Synt)-1A knock-out cells show impaired docking and 1st phase, but not 2nd phase, secretion (Ohara-Imaizumi et al., J Cell Biol 2007)

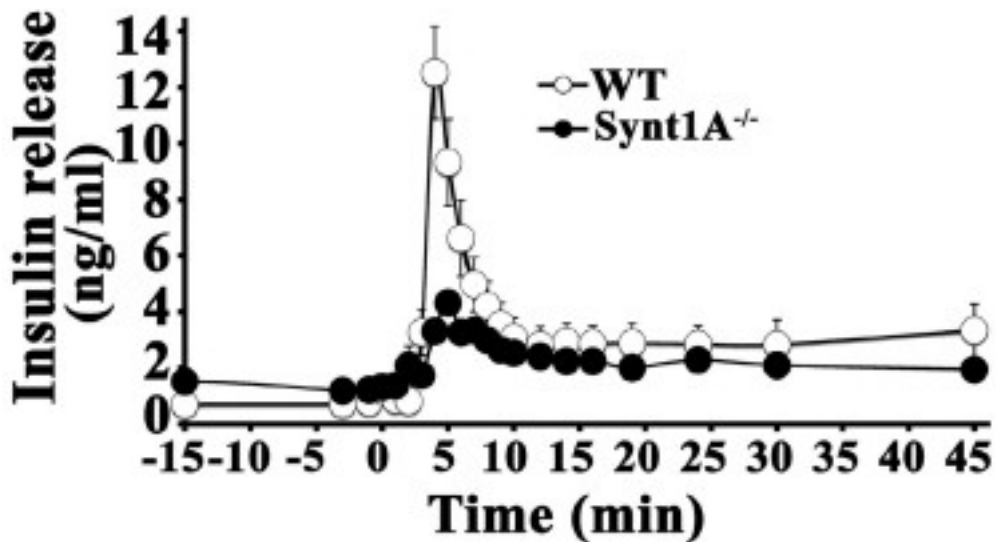
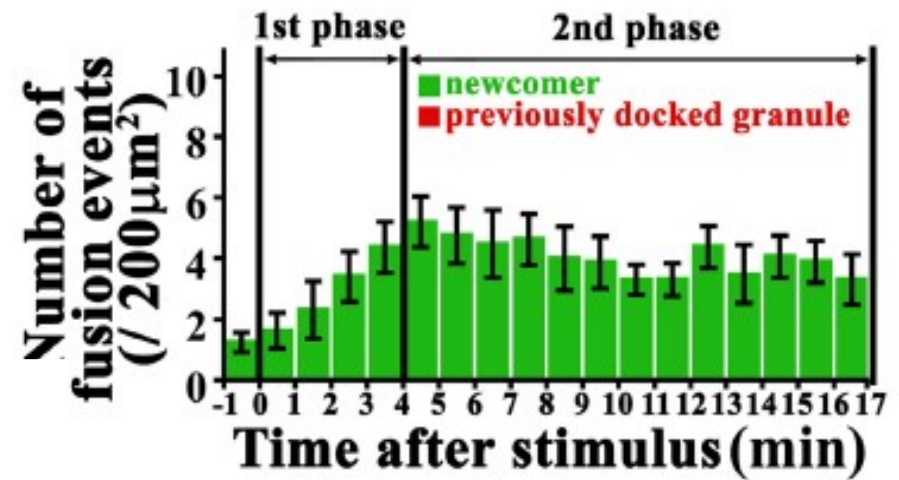


Due to fusion of newcomer granules

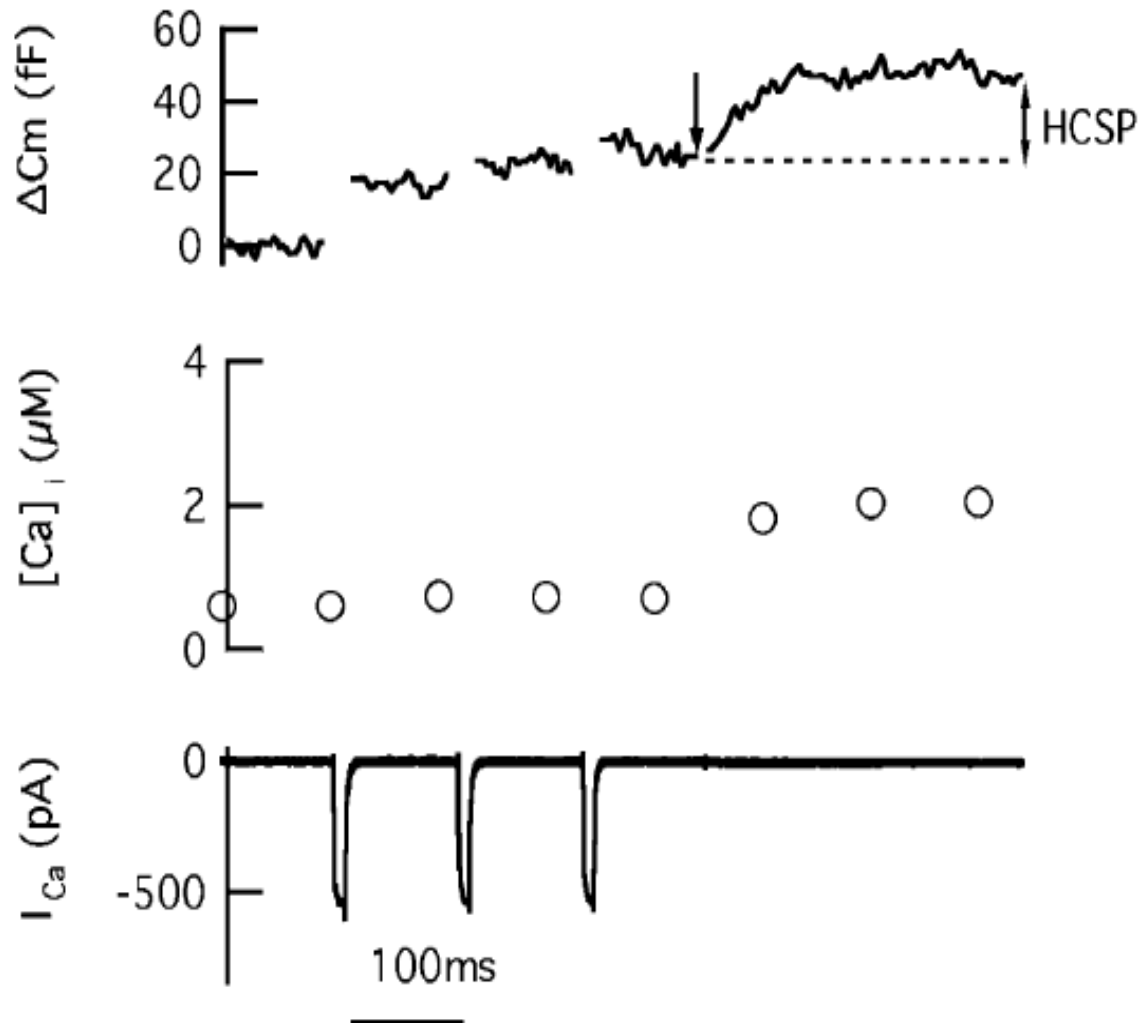
WT



KO



Highly calcium sensitive pool (HCSP) of granules

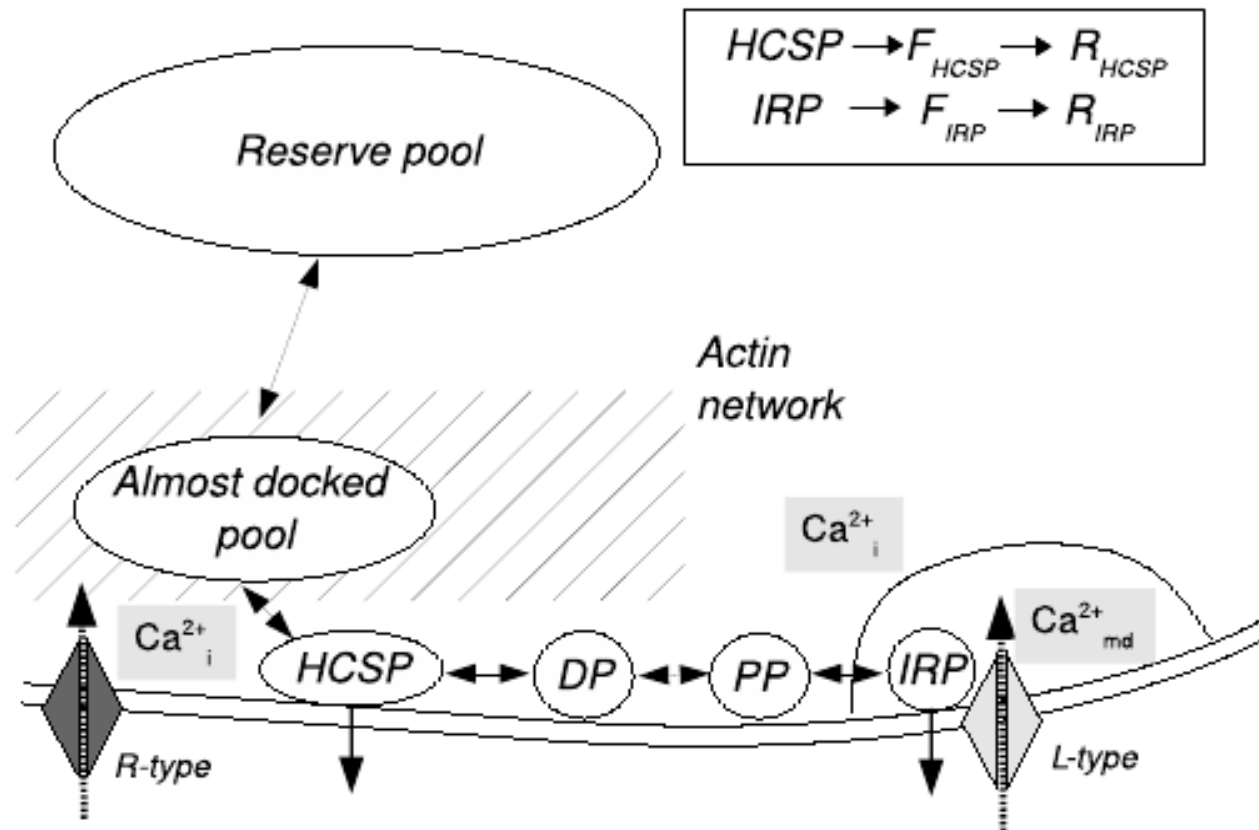


- Wan et al. + Yang & Gillis (JGP 2004)
- Affinity $\sim 2 \mu M$
($\sim 20 \mu M$ for RRP)
- The HCSP resides away from calcium channels since depolarizations do not empty it
- Newcomers also fuse away from calcium channels

Including the HCSP

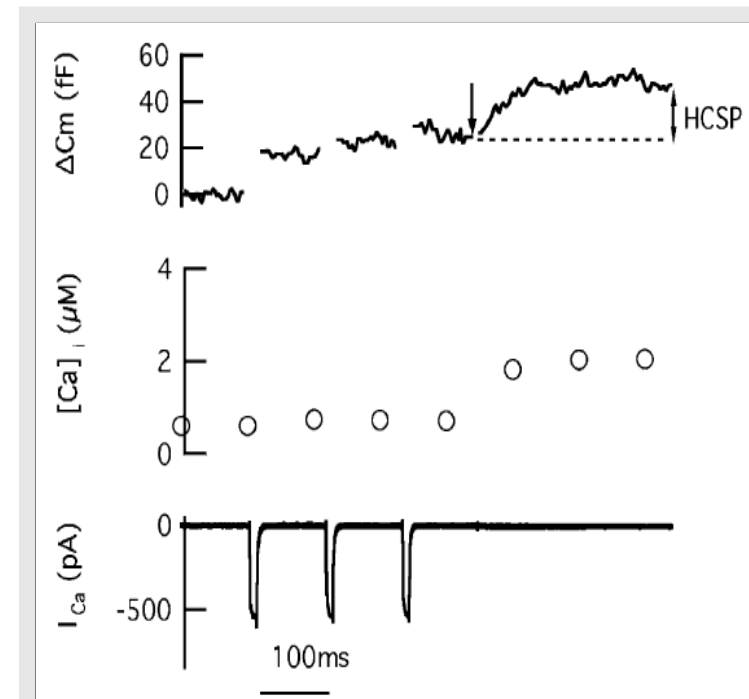
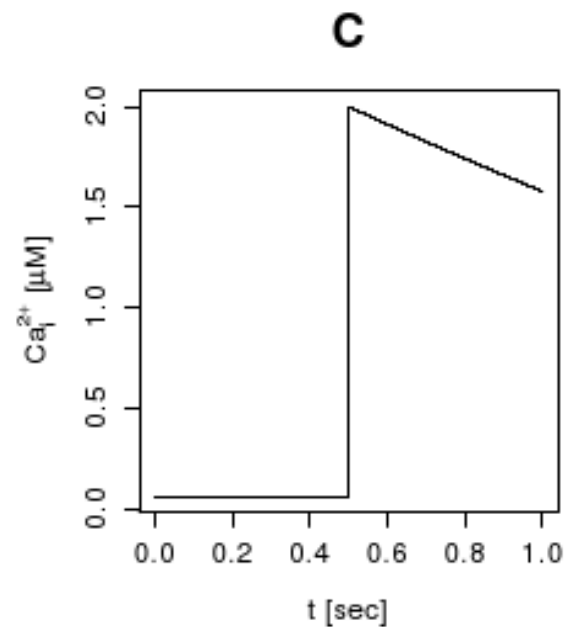
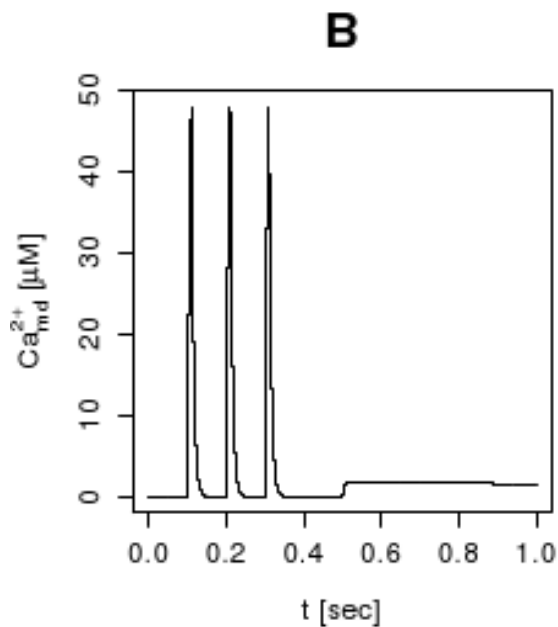
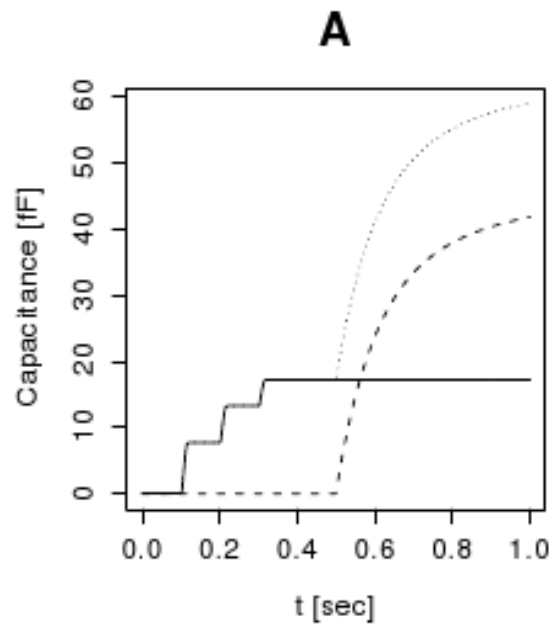
... in the model by Chen et al. (Biophys J 2008)

- Distinction between global, cytosolic and local, microdomain calcium, and between L- and R-type calcium channels
- HCSP assumed to reside away from Ca^{2+} channels
- HCSP assumed to be independent of syntaxin-1A, and to consist of granules that are tethered, but still not completely docked

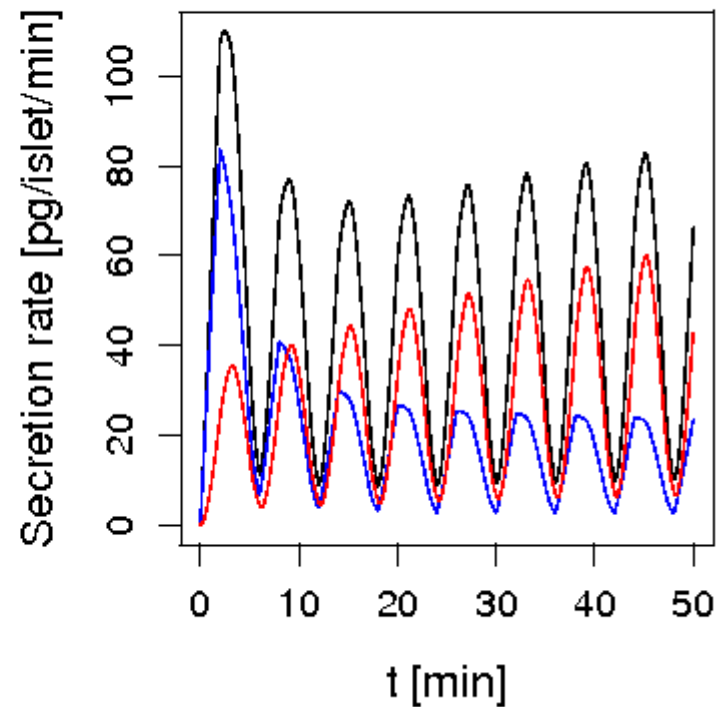
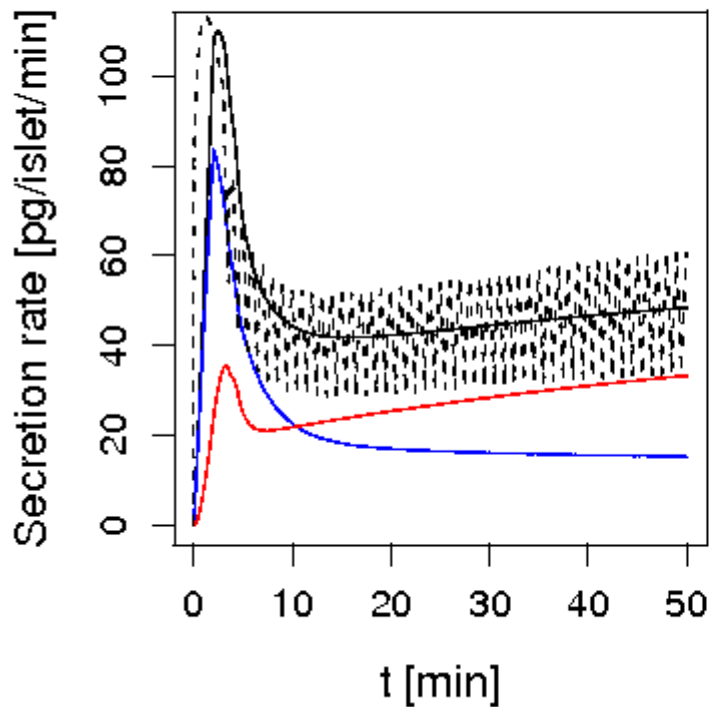


Pedersen & Sherman (PNAS 2009)

Simulations: Yang & Gillis protocol

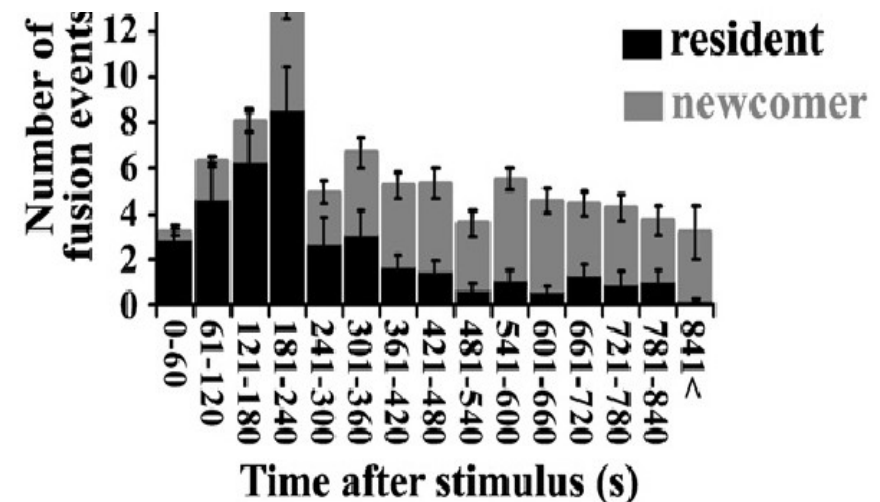
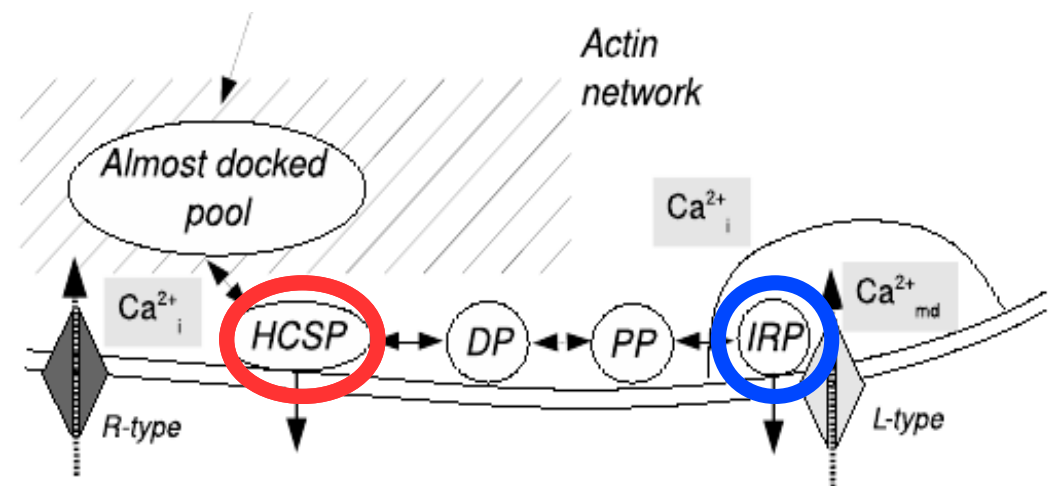


Simulations: Newcomer granules fuse from the HCSP



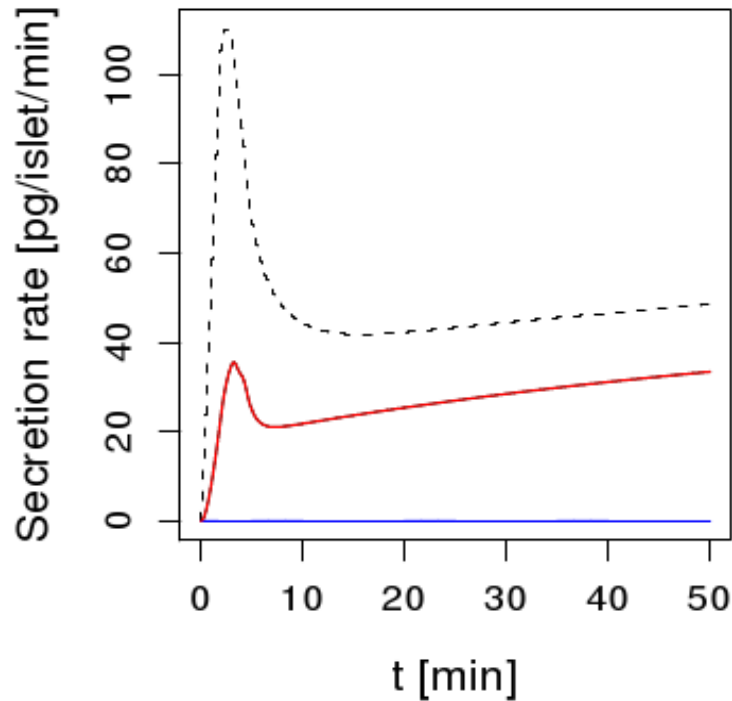
IRP secretion
HCSP secretion
Total secretion

2nd phase →

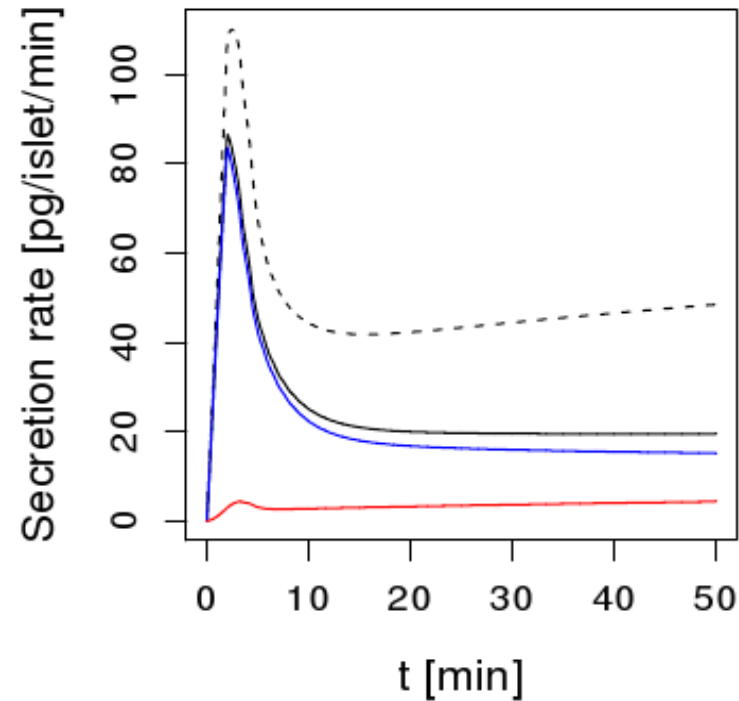


Simulations: Calcium channel KO

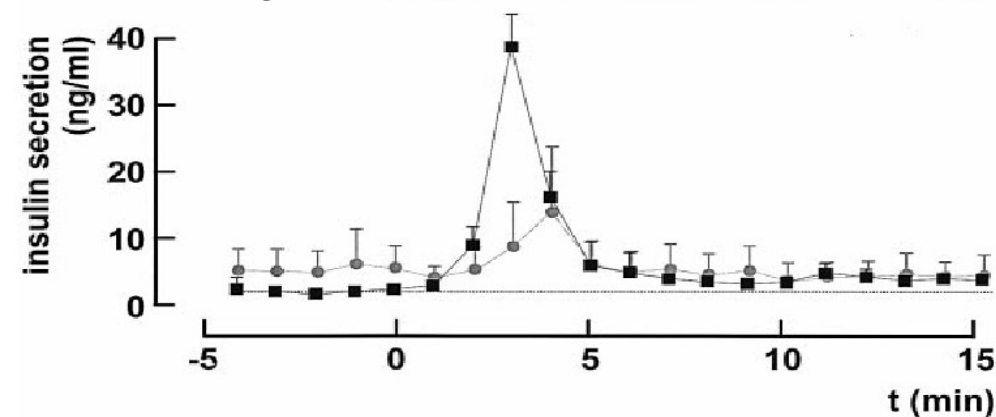
L-type KO/block
(Schulla et al., EMBO J 2003)



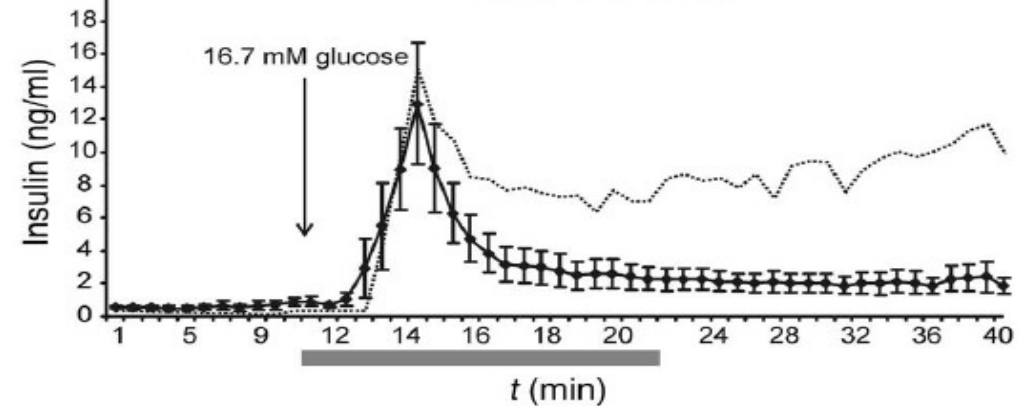
R-type KO/block
(Jing et al., JCI 2005)



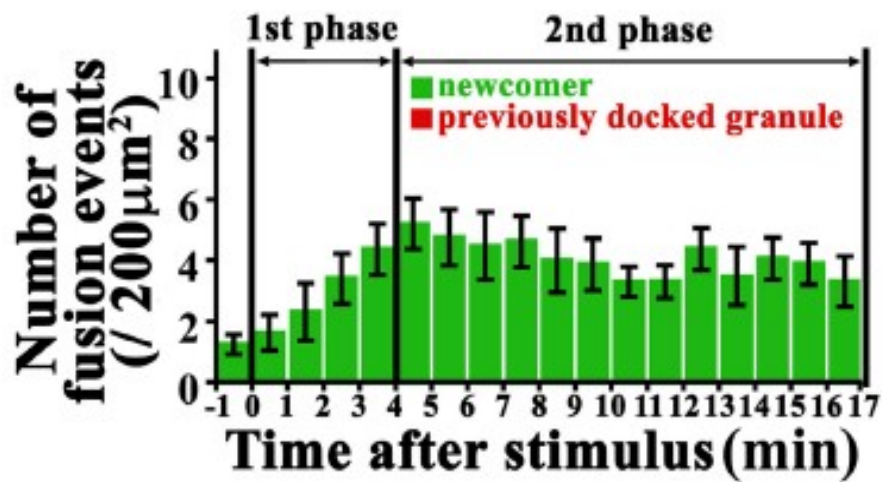
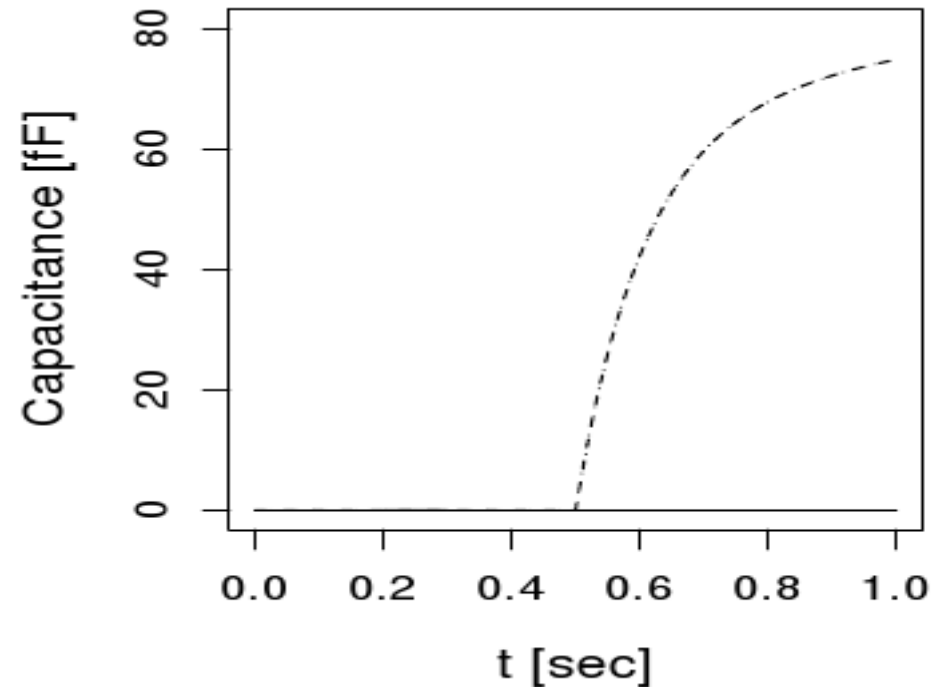
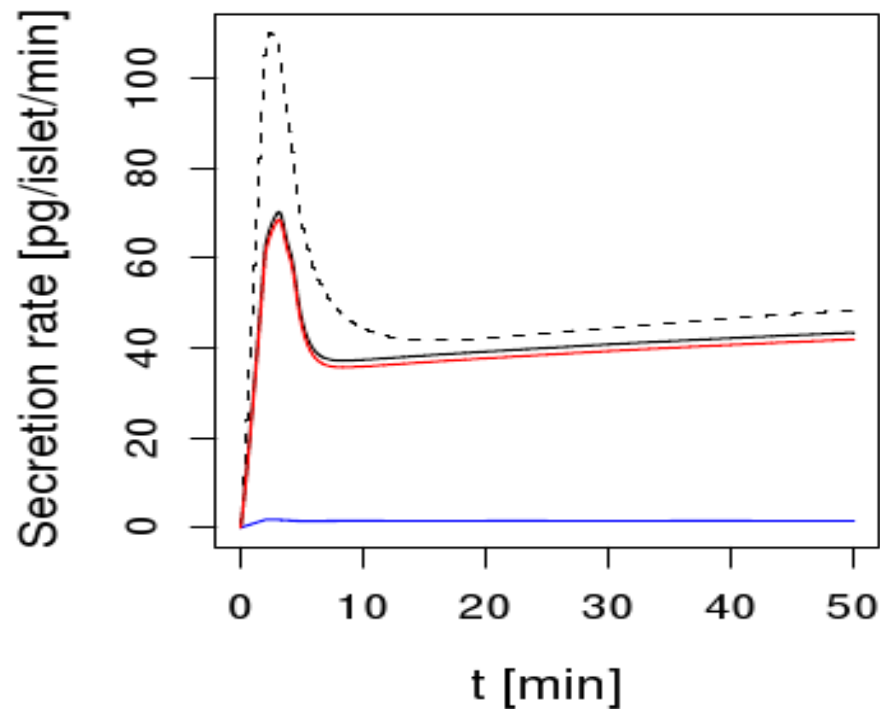
10 mM gluc



D NMRI + SNX482



Simulations: Synt1A KO cells by assuming reduced docking rate



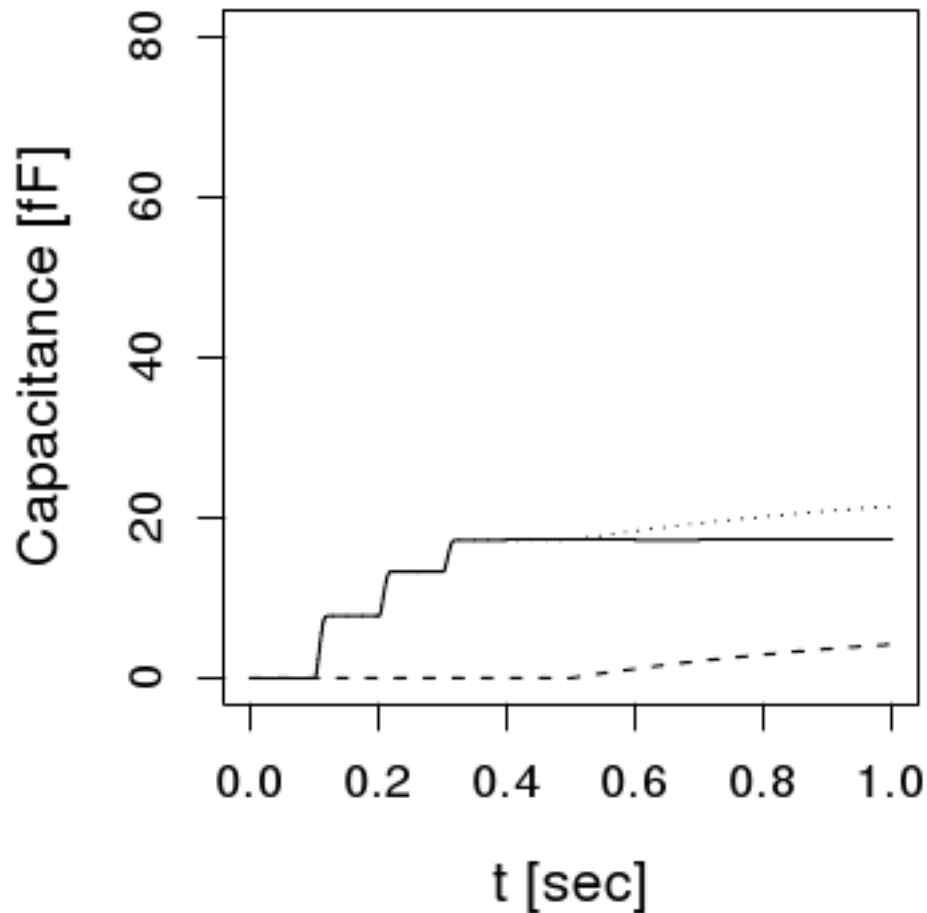
Prediction and crucial test of the model:

The HCSP is intact and possibly increased in Synt1A cells

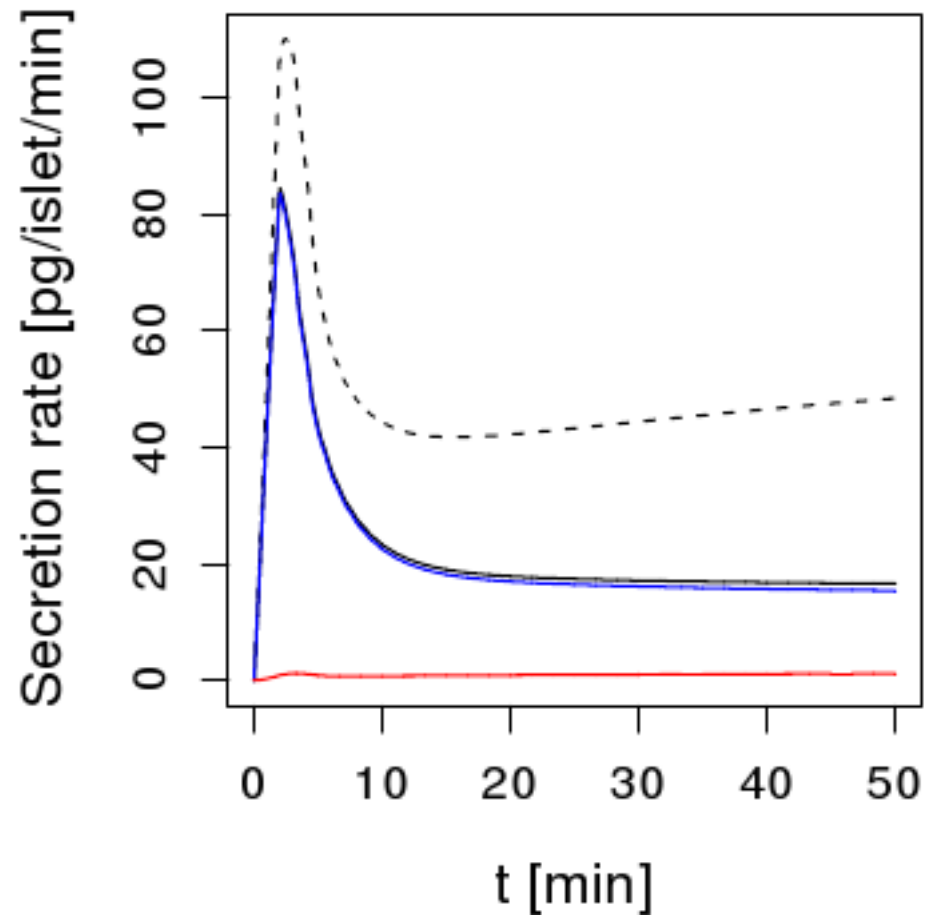
Different calcium sensors?

- Synaptotagmins (Syt's) are believed to be the sensors of calcium
 - Syt-9 is a low affinity (tens of μM) sensor present in beta-cells, and is likely the IRP sensor
 - Syt-7 and Syt-3 are high affinity (few μM) sensors, and have been suggested to be involved in insulin secretion (Syt-3 controversial). Could be the HCSP sensors

Simulations: KO of the HCSP sensor

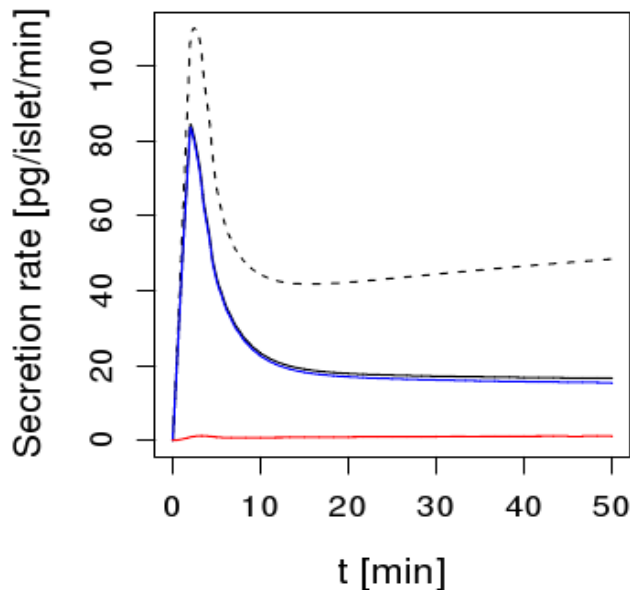
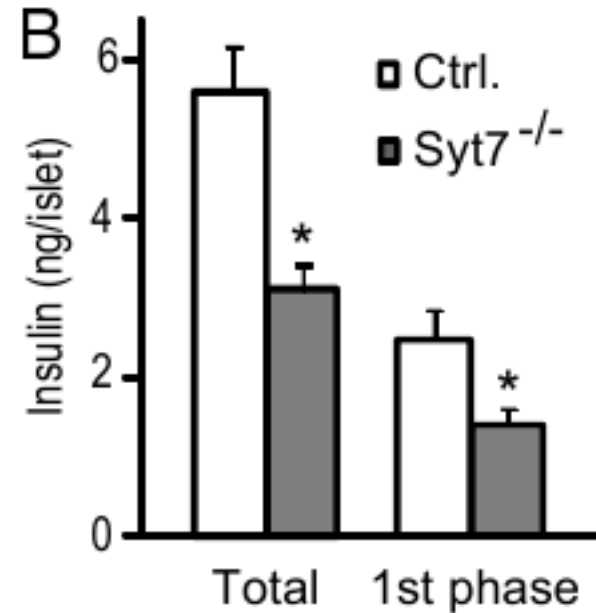
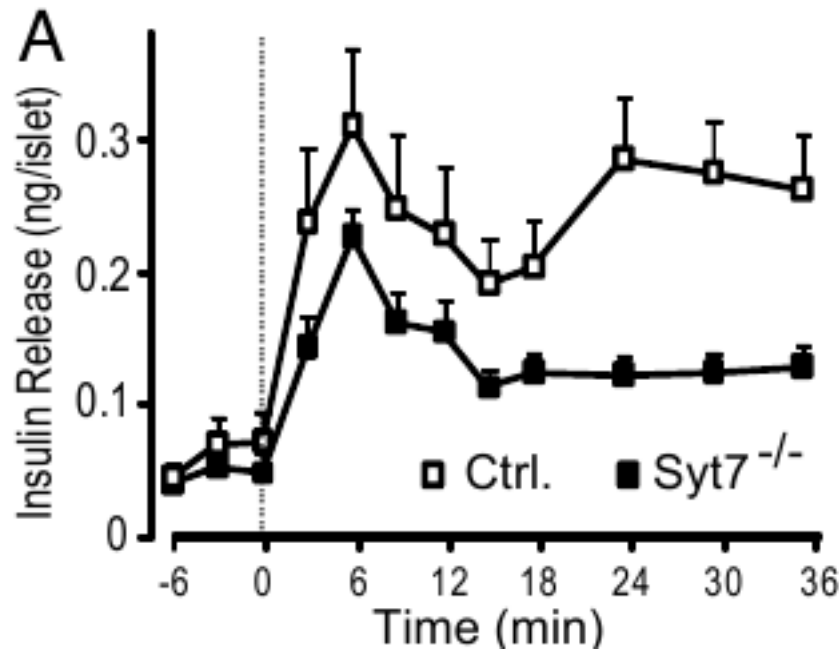


... as expected



Prediction: Second phase secretion is impaired

Is Syt-7 the HCSP sensor?



Gustavsson et al. (PNAS 2008)

Both reduced 1st and 2nd phase
– but there might be Syt-7/Syt-9
interactions changing the properties of
IRP release (Schonn et al., PNAS 2008)

Summary (part 2)

- Part 1 bridges levels of organization (*granules* → *organ*)
- Part 2 spans timescales by coupling secretion (*minutes*) to capacitance measurements (*milliseconds*) for various perturbed situations
 - Ca^{2+} channels KO/blocking
 - Syntaxin-1A KO
 - Ca^{2+} sensor/synaptotagmin KO
- Including a HCSP as a transient state away from L-type calcium channels, naturally identified the HCSP with newcomer granules.
- Mathematical modeling was used to test the plausibility of the biological hypothesis

Conclusions

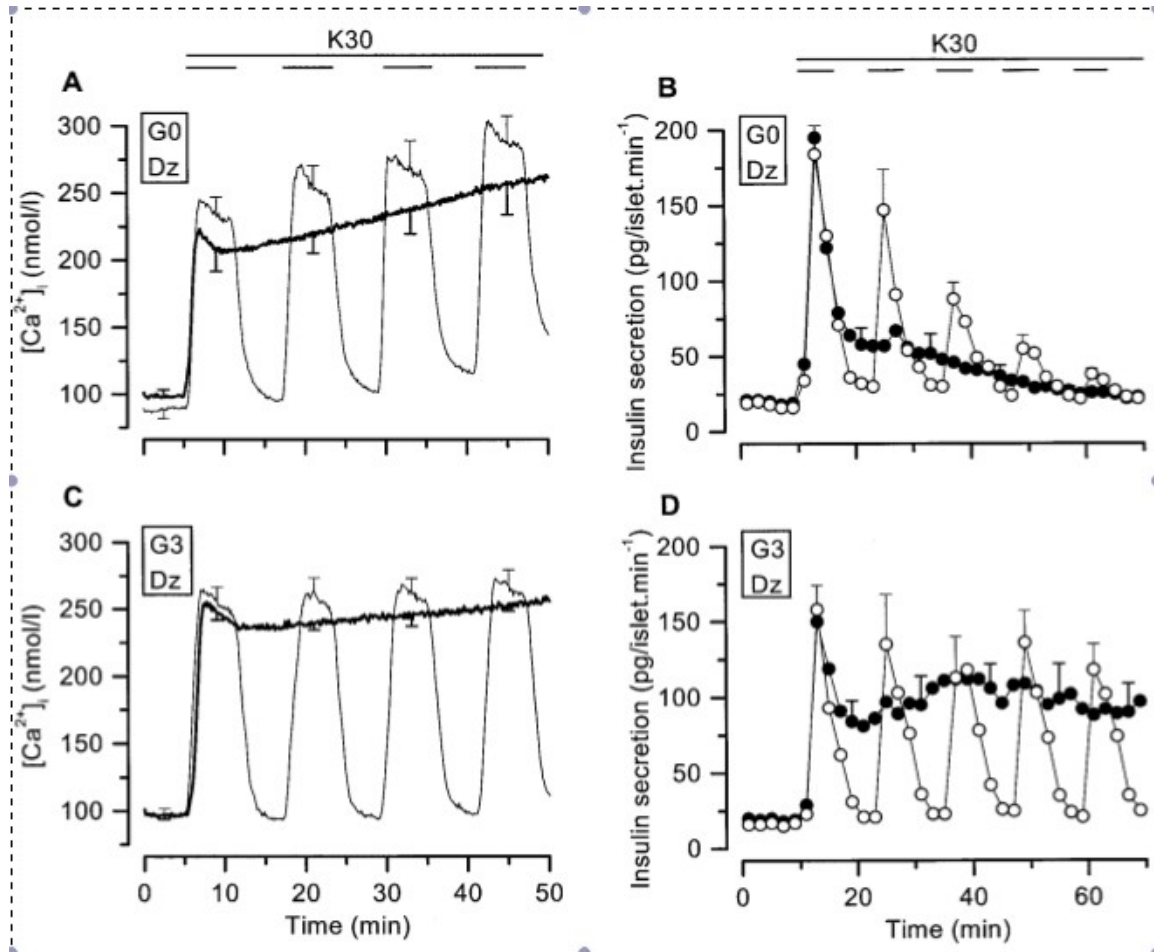
- Mathematical models are used to integrate separate experiments in a structured, coherent way
- Can be used to span timescales and levels of organization
- Two classes:
 - “Models to simulate”
(test hypotheses, predict outcome of experiments; can include different levels of detail depending on the scope of the model)
 - “Models to measure”
(extract information from data; must be simple/minimal to allow parameter estimation)

Conclusions

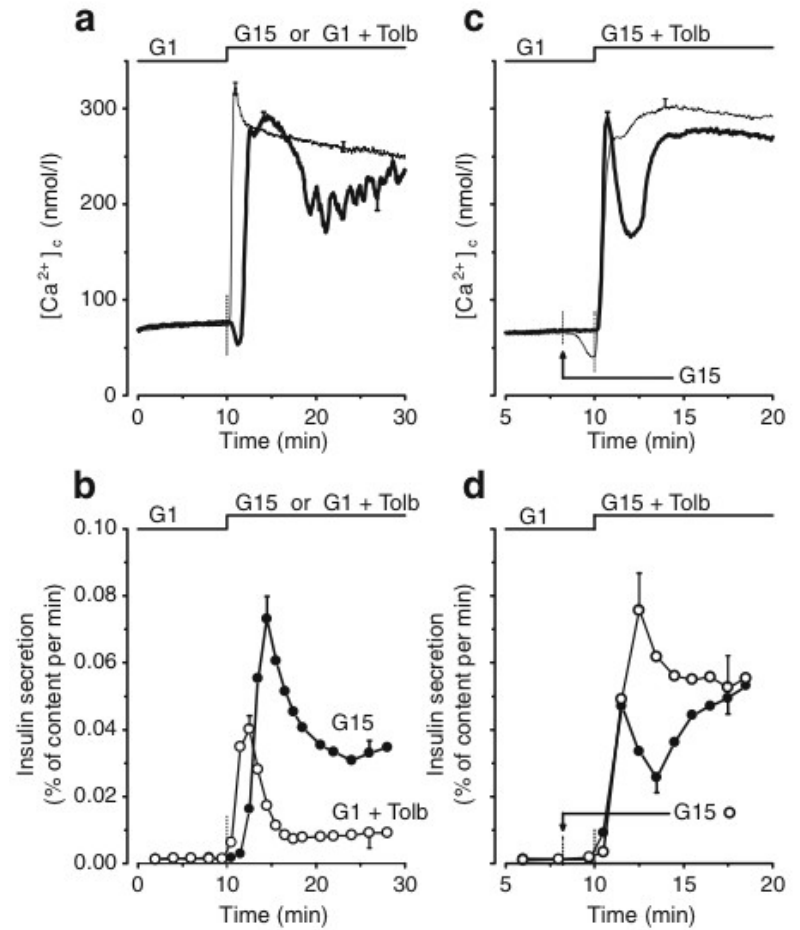
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Thank you!

Pools?



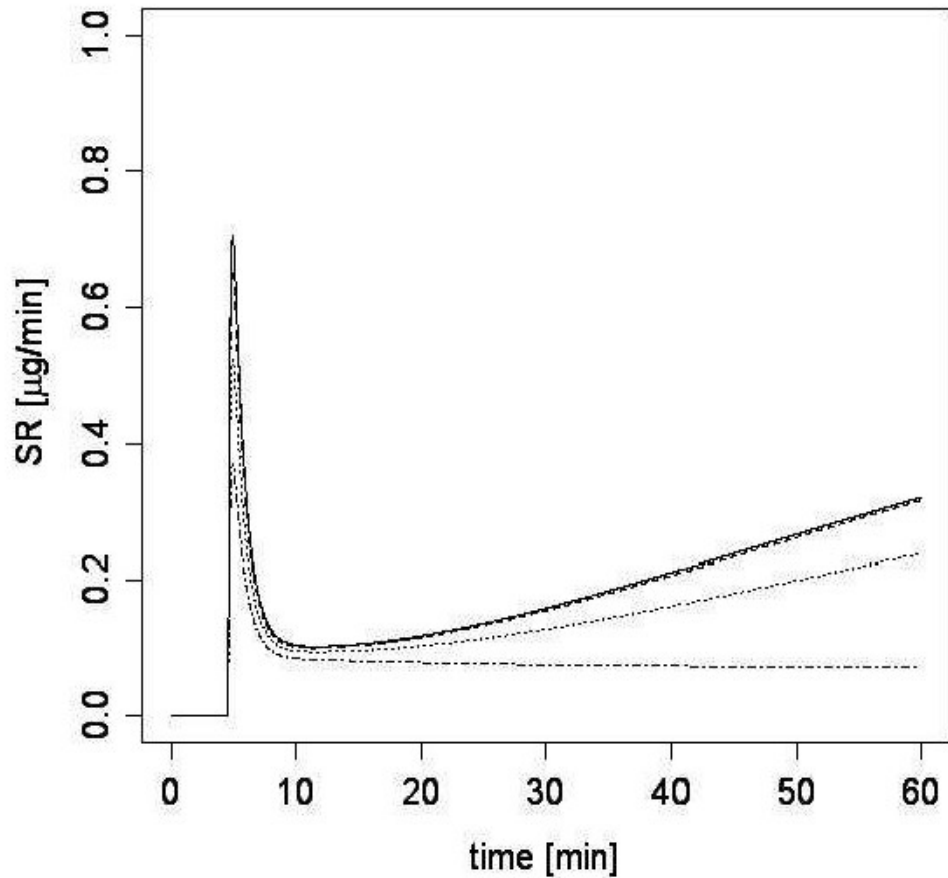
Henquin, 2002



Henquin, 2009

Results

Biphasic



Potentialiation

